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## **Cross-Species *C. Elegans* Model of Opioid-Induced Behavior Identifies Gao and a Multi-Layered RGS Profile Mediating Opioid Receptor Signaling**

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We previously pioneered a cross-species transgenic model of mu-opioid receptor signaling (tgMOR) using *C. elegans*. In the tgMOR model, mammalian MOR is expressed in the *C. elegans* nervous system imbuing the animal with robust opioid-responsive behaviors. Previous unbiased, forward genetic studies with the tgMOR model revealed novel, conserved regulators of MOR signaling that influence opioid responses from *C. elegans* through rodents. At present, we know little about how tgMOR engages endogenous neuronal signaling components to generate opioid-responsive behaviors. Here, we investigate how G-protein signaling and RGS proteins influence opioid sensitivity. Forward genetics identified tgMOR; goa-1 mutants as opioid hypersensitive. GOA-1 is the sole conserved G $\alpha$ o protein in *C. elegans*. CRISPR engineered null and deletion mutants for tgMOR; goa-1 displayed impaired behavioral responses to fentanyl. To further investigate tgMOR signaling, we CRISPR engineered early stop cassettes into two evolutionarily conserved RGS proteins, eat-16 (mammalian RGS9) and egl-10 (RGS6/7). Our results demonstrate both tgMOR; eat-16 and tgMOR; egl-10 mutants are hypersensitive to fentanyl. Our findings further indicate RSBP-1 (R7BP) and Gq signaling influence opioid sensitivity. Overall, our observations support two key conclusions: 1) G $\alpha$ o is potentially an ancient evolutionarily conserved mediator of MOR signaling. 2) Engineered MOR signaling in *C. elegans* creates a multi-layered RGS regulatory profile that restricts tgMOR/GOA-1 signaling with EGL-10 being a primary inhibitor and EAT-16 playing a secondary inhibitory role. Ongoing large-scale genetic studies with the tgMOR *C. elegans* model are now poised to reveal further genetic targets for potentially managing opioid substance use disorder.