Name: Schahram Akbarian Email: schahram.akbarian@mssm.edu

Chromatin Fiber Profiling and Transcription Factor Footprinting by Genome-Scale Single Molecule Sequencing from Opioid Use Disorder and Control Brain

Aman Agarwal¹, Bicheng Jiang¹, Cyril J Peter¹, Risa Watanabe¹, Deborah Mash³, Schahram Akbarian^{1,2,4}

Department of Psychiatry, Icahn School of Medicine at Mount Sinai, New York, NY 10029
Department of Neuroscience, Friedman Brain Institute, Icahn School of Medicine at Mount Sinai, New York, NY 10029

 ³Nova Southeastern University, Fort Lauderdale FL 33328
⁴Department of Genetics and Genomic Sciences, Center for Advanced Genomics Technology, Icahn School of Medicine at Mount Sinai, New York, NY 10029

We established in our laboratory a Fiber-seq protocol for the human brain, which allows for the first time to profile single chromatin fibers, directly from brain nuclei, uninterrupted for 10-20kb on a genome-wide scale without the need for PCR or other amplification. This type of single molecule long-read sequencing provides, at base pair resolution, information on m5CpG methylation patterns, nucleosomal positioning and the activity-status of nucleosome depleted regions by transcription factor footprinting. Furthermore, we bypass additional limitations posed by conventional nucleolytic assays (incl. ATAC-seq) which typically built chromatin landscapes from short-read sequencing (0.1-0.15kb) of PCR amplified DNA, which leaves an estimated 50% of human nuclear genome underexplored because of poor annotation of low-complexity repetitive loci, duplicated regions, tandem arrays, and complex structures.

Here, we apply, for the first time, our fiber-seq protocol to the frontal cortex of a cohort of opioid use disorder and control brains. We will present initial results on single fiber-level epigenomic dysregulation in diseased brains, with multiomic integration of endogenous CpG methylation, and nucleosomal positioning at promoter and enhancer regions including potential changes in transcription factor footprints.

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