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Molecular characterization of cocaine-seeking engram neurons in the infralimbic cortex using single nuclei RNA-sequencing in rats

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The risk of relapse is a cardinal feature of addiction. Environmental stimuli previously paired with drug-taking can elicit drug seeking and precipitate relapse long after cessation of drug use. One strategy to combat relapse is to inactivate or weaken these persistent maladaptive drug-cue associative memories by targeting the neurons encoding this memory representation (engram). Engram neurons are commonly identified by their transient expression of immediate early genes (such as *Fos*) after the association experience, and Fos-expressing engram neurons play a causal role in drug-seeking behaviors. However, the molecular mechanisms and cell types that reinforce this drug-cue association are unknown. We hypothesized that the drug-cue engram in association cortices comprises multiple cell types with unique transcriptional signatures to support future cue-induced drug seeking.

Here we use single nuclei RNA sequencing and a Fos-based transgenic rat (Fos-mRFP) to characterize drug-cue engram neurons in the infralimbic cortex. We trained rats to self-administer cocaine during twice daily 3 h sessions. Following 21 days of abstinence, we tested rats for cocaine-seeking (30 min, extinction conditions) and collected brains 3 h after test (peak Fos-driven mRFP expression). We used fluorescence-activated nuclei sorting to isolate mRFP-positive (engram) and mRFP-negative (non-engram) nuclei as input for snRNA-seq. Using this unbiased approach, we characterize cell-type and engram-specific transcriptional signatures that contribute to drug-seeking behaviors. We will employ CRISPR-based transcriptional modulators in future experiments to assess causal roles for these cocaine memory-specific genes in relapse.